

Mechanisms of cortisol-mediated inhibition of reproductive hormones in heat stressed cows: A mini-review

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Abstract

The reproductive performance of cows is significantly influenced by stress-induced cortisol release, especially in situations of physiological and environmental stress, like heat stress. The zona fasciculata of the adrenal cortex produces cortisol, a glucocorticoid hormone that is a major indicator of stress, and is mostly controlled by the hypothalamic-pituitary-adrenal (HPA) axis. The processes by which high cortisol levels negatively impact bovine reproduction are examined in this review. Increased cortisol concentration in the circulatory system impairs follicular development, ovulation and oestrus behaviour by blocking the secretion of gonadotropin-releasing hormone (GnRH) and gonadotrophins [luteinizing hormone (LH) and follicle stimulating hormone (FSH)], which upset the hypothalamic-pituitary-gonadal (HPG) axis. Cortisol affects steroidogenesis, lowers oocyte quality and modifies folliculogenesis at the ovarian level by interacting with glucocorticoid receptors. Furthermore, the important reproductive hormones like progesterone and oestradiol are suppressed by stress-induced cortisol which results in poor follicular maturation, decreased embryo viability and lower conception rates. Long-term stress especially heat stress, increases embryonic mortality and modifies gene expression linked to embryonic development thereby aggravating these consequences. In general, high cortisol levels negatively affect fertility by disrupting oocyte competence, hormonal balance and gestational processes. Developing efficient management techniques to reduce stress and increase the reproductive performance of cows requires an understanding of these mechanisms.

Keywords: Cattle (cows); Heat stress; Cortisol; Female reproduction; Folliculogenesis; Gonadotropin-releasing hormone.

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Introduction

Cortisol ($C_{21}H_{30}O_5$), which is secreted by the zona fasciculata of the adrenal cortex of the adrenal gland, is the hormone that is mostly produced during stress, and is a helpful biochemical marker of stress in animals (Martinez *et al.*, 2016). According to Massey *et al.*, (2014), cortisol is a glucocorticoid hormone that has been shown to be a reliable biological correlate of a number of detrimental health consequences. It is involved in various processes such as metabolism, blood pressure and immune response control.

When exposed to acute heat stress, plasma cortisol levels rise within twenty minutes and plateau in two hours (Christison and Johnson, 1972). Since higher cortisol concentrations may be associated with higher levels of stress and pain, cortisol is the hormone most frequently employed to identify and quantify the extent of the stress and pain response (Molony *et al.*, 1993; Fisher *et al.*, 1996; Ley *et al.*, 1996). However, animals that have lower cortisol concentrations are unable to experience stress (Stilwell, 2012). Cortisol is one of the primary hormones generated during stress reactions in animals because stress can activate the hypothalamic-pituitary-adrenal system, and it is easily activated with increasing cortisol production (Kalin *et al.*, 1992).

There is clear evidence that hyperthermia is detrimental to all forms of production from the perspective of animal welfare at both high ambient temperatures and high direct and indirect solar radiation (Silanikove, 2000). In cattle, heat stress results in ovarian inactivity due to hyper-prolactinemia, decreased luteinizing hormone frequency, inadequate follicle maturation and decreased oestradiol synthesis (Wolfenson *et al.*, 2000). Adrenocorticotrophic hormone (ACTH) may be involved in some symptoms of heat stress. Additionally, high temperatures disrupt the

release of some reproductive hormones in cows such as progesterone, follicle-stimulating hormone (FSH) (Howell *et al.*, 1994; Burke *et al.*, 2001; Wise *et al.*, 1988; Gilad *et al.*, 1993). Heat stress reduces follicular steroidogenesis, androstenedione and oestradiol levels in the follicle wall, according to an in vitro study from bovine species (Bridges *et al.*, 2005).

Mechanism of action of stress-induced cortisol inhibition of reproductive performances via endocrine interactions

Stress triggers the hypothalamic-pituitary-adrenal (HPA) axis (Minton, 1994). The interactions between HPA axis with the hypothalamic-pituitary-gonadal (HPG) axis influence the reproductive system negatively (Collier *et al.*, 2017). HPA axis activation causes the secretion of corticotropin releasing hormone (CRH) into the median eminence by neurones in the hypothalamus. Adrenocorticotrophic hormone (ACTH) is released from pituitary corticotroph cells when CRH passes through the hypothalamic-pituitary portal system after secretion from the neurons of the hypothalamus. The adrenal gland produces and secretes glucocorticoids (cortisol) in response to adrenocorticotrophic hormone stimulation. Stressors that activate the HPA axis may result in infertility by preventing LH secretion (Breen and Karsch, 2006).

Gonadotropin releasing hormone (GnRH) neurones' cell bodies are located where certain CRH neurones in the hypothalamus end (Wade and Jones, 2004). GnRH release from GnRH neurones may be inhibited when CRH neurones are activated and release CRH. Because GnRH neurones lack the type II glucocorticoid receptor, glucocorticoids have inhibitory effects on the release of GnRH and LH, but these effects are not directly mediated by glucocorticoids. The hypothalamic kisspeptin, neurokinin B and dynorphin (KNDy) cells have a glucocorticoid receptor and can

provide the glucocorticoid signal to the GnRH neurones (Ralph *et al.*, 2016; Scott *et al.*, 2019).

Long-term stressors, like heat stress, typically trigger the sympatho-adrenal system and the hypothalamo-pituitary-adrenal axis (Tibrook *et al.*, 2000). Corticotrophin-releasing factor (CRF) and arginine vasopressin (AVP) neurones in the paraventricular nucleus are activated when the hypothalamo-pituitary-adrenal axis is stimulated and these neuropeptides are secreted into the hypophyseal portal system to stimulate the corticotrophs of the anterior pituitary gland (Tibrook *et al.*, 2000). Heat stress causes the corticotrophs to secrete melanocyte-stimulating hormone, endorphin and ACTH among other peptides generated from pro-opiomelanocortin (Engler *et al.*, 1989). ACTH acts on the cortex of the adrenal glands to boost the synthesis and production of glucocorticoids like cortisol, however the physiological significance of the release of endorphin and melanocyte-stimulating hormone in response to heat stress is unclear. Heat-stressed cows may produce higher concentrations of this catabolic hormone to sustain milk supply (Albiley *et al.*, 1975).

The triggering of the hypothalamic-pituitary-adrenal (HPA) axis due to stress reduces livestock species' ability to reproduce, causing infertility (Collier *et al.*, 2017). The pituitary gland releases adrenocorticotrophic hormone (ACTH) which triggers the release of catecholamines and glucocorticoids or cortisol, both of which significantly act to cushion the effects of stress and have inhibitory effect on reproduction. Heat stress is linked to the persistent release of ACTH which causes the adrenal gland to produce cortisol which affects ovulation and follicular growth by changing the effectiveness of follicular selection and dominance.

Effects of cortisol and glucocorticoid activity on the function of the bovine ovary (cumulus cell) during ovulation

Although the ovaries lack the enzymes required for cortisol manufacture and they do exhibit glucocorticoid receptors, they are impacted by cortisol. The glucocorticoid receptor (GR) is a receptor through which cortisol works (Coutinho and Chapman, 2011; Hapgood *et al.*, 2016; Timmermans *et al.*, 2019). Regardless of their level of maturity, bovine cumulus cells have been demonstrated to express GR (Anbo *et al.*, 2022). In the development of cumulus oophorus, local cortisol synthesis increases concurrently with progesterone (P₄) production (Tetsuka *et al.*, 2016; Tetsuka and Tanakadate, 2019). The steroidogenic enzyme 11 β -hydroxysteroid dehydrogenase type1 (HSD11B1) mediates this process by converting the inert glucocorticoid cortisone into active cortisol (Krozowski *et al.*, 1999; Michael *et al.*, 2003; Seckl, 2004; Holmes and Seckl, 2006). Additionally, the locally synthesised P₄ in the cumulus cells increases the expression of HSD11B1 and cortisol production (Anbo *et al.*, 2022). These findings imply that glucocorticoids are produced and function in bovine cumulus cells, and that P₄ can activate this system during the periovulatory phase. When follicular synthesis of progesterone and 17 α -hydroxyprogesterone reaches levels that displace cortisol from its binding proteins and make it available for biological action, ovarian follicles then regulate the biological activity of cortisol. Free cortisol, which is present in the pre-ovulatory follicle soon before ovulation, works to lessen the inflammatory-like processes that accompany ovulation. This explains why the presence of a dominant follicle during oestrus is associated with rise in serum levels of cortisol (Andersen, 2002; Ram *et al.*, 2023).

Effects of stress-induced cortisol on folliculogenesis, oocytes and ovulation

Folliculogenesis is controlled at the endocrine level by a cascade mechanism involving the ovary, anterior pituitary and central nervous system. Gonadotropin-releasing hormone (GnRH) is secreted into the portal blood vessels by specialized hypothalamic neurons, which act on gonadotrophs to cause a pulsatile release of follicle-stimulating hormone (FSH) and luteinizing hormone (LH), which in turn act on ovarian follicle cells to regulate folliculogenesis. Similarly, follicle derived hormones and growth factors can act locally to modify (amplify or attenuate) the activity of GnRH, FSH and LH which are crucial in controlling folliculogenesis (Kumar and Sharma, 2014; Hsueh *et al.*, 2000). When high level of cortisol is sustained throughout the late luteal and early follicular phases of the oestrous cycle, there is inhibition of follicular growth and development, which prevents the preovulatory surge of LH (Doney *et al.*, 1973; Welsh and Johnson, 1981; Wilson *et al.*, 1998). Corticosterone alters the molecular pathways underlying oocyte maturation and competence for future development by reducing phosphorylated forms of extracellular signal-regulated kinases (ERKs), phospho-ERK (p-ERK)-1 and p-ERK-2 in exposed oocytes. This represents a change in the molecular pathways driving oocyte maturation and competence for later development (Gonzalez *et al.*, 2010a).

Heat stress genes like HASP14 and Cx34 become active when the outdoor temperature climbs above 25°C in summer. Since these stress genes are linked to apoptosis among other processes, apoptosis can be important for both the development of the embryo and the follicle/oocyte from the start to metaphase II (Pavani *et al.*, 2016; 2017). Insufficient gonadotropin production is the cause of follicular development and atresia during anestrus (Das and Khan, 2010).

According to Ranabir and Reetu (2011), cortisol has an adverse influence on oocyte maturation and subsequent embryonic development following IVF. Exposing oocytes to high concentrations of cortisol reduced their competence and impaired oocyte developmental potential by causing ovarian cells to undergo apoptosis through the activation of a transmembrane protein which is part of the tumour necrosis factor (TNF) and is commonly referred to as Fas ligand (FasL). Studies conducted on cortisol-injected mice showed that cortisol has a definite detrimental effect on oocyte nuclear maturation rate (NMR), blastocyst rates and the number of cells per blastocyst (Raghavendra *et al.*, 2007; Yuan *et al.*, 2016).

Effects of heat stress induced rise in cortisol levels on reproductive hormones

Glucocorticoids play a crucial role in mediating the inhibitory effect on reproduction during heat stress by directly preventing oocyte meiotic maturation. Additionally, corticotropic releasing hormone (CRH) inhibits ovarian steroidogenesis which is caused by a decrease in luteinizing hormone (LH) secretion (Al-Katanani *et al.*, 2002). ACTH has been shown to prevent estradiol-induced sexual behaviour (Heinz and Allrich, 1992) and heat stress can enhance cortisol release (Roman-Ponce *et al.*, 1981; Wise *et al.*, 1988; Elvinger *et al.*, 1992). Since this can inhibit GnRH and hence LH secretion, increased corticosteroid secretion has been proposed (Gilad *et al.*, 1993).

Heat stress may inhibit oocyte growth by reducing the synthesis of pre-ovulatory surges in luteinizing hormone and oestradiol, which results in poor follicular maturation and ovarian inactivity in cattle (Hansen, 2007). High cortisol levels can affect the physiology of reproduction because it decreases the amplitude of GnRH and LH secretion as well as the levels of plasma oestradiol in the follicular phase (Wagenmaker *et al.*, 2009; De Graaf

Roelfsema *et al.*, 2007). According to Lotfi and Mendonca (2016) and Lee *et al.*, (2007), cortisol suppression of the protein ERKs disrupts their functions in meiotic maturation of fully developed oocytes and/or arrest at metaphase of meiosis II before fertilization. Numerous studies suggest that glucocorticoids in general and cortisol in particular may contribute to the anti-gonadal action of stress, even if the exact causal relationship between stress and infertility has not been established (Dobson and Smith, 1995; Chrousos *et al.*, 1998). Furthermore, elevated cortisol levels decrease the generation of oestradiol by potentially altering granulosa cell activities within the follicle leading to a decline in oocyte quality and a reduced capacity to develop following fertilization (Prasad *et al.*, 2016).

Effects of heat stress-induced cortisol on bovine fertility and reproductive efficiency

Several *in vivo* studies have shown that rise in cortisol levels specifically impacts bovine metabolism and reproduction (Mudron *et al.*, 2005; Pavani *et al.*, 2016). Extreme weather, travel, laparoscopy and psychological stress are examples of stressful stimuli that suppress or decrease the manifestation of behavioural oestrus and ovulation which lowers fertility in domestic species. These stressors do not only lower fertility but also activate the hypothalamic-pituitary-adrenal (HPA) axis. Stressors related to management are frequently linked to a significant rise in cortisol levels in the blood (Prasad *et al.*, 2016; Mahdy *et al.*, 2017; Komesaroff *et al.*, 1998).

Earlier reports show that bovine fertility sharply declines to 35% during the summer and reaches 78% during the cooler seasons of spring and autumn (Pavani *et al.*, 2017). A number of theories have been proposed to explain this closure during warmer seasons (Mahdy *et al.*, 2017; Pavani *et al.*, 2016; 2017). When calves are subjected to stressors like

high temperatures for extended periods of time, cortisol levels can rise to a maximum of approximately 250 μ M (Polsky and Von Keyserlingk, 2017; Gonzalez, 2010b). According to Pavani *et al.*, (2015), the effects of high ambient temperatures on reproductive processes include decreased bovine fertility throughout the summer, altered physiological mechanisms, inhibition of *in vitro* oocyte maturation and low *in vitro* embryonic development. When cows were exposed to a mild heat stress (35°C) over an extended period of time, their plasma cortisol levels dramatically increased (Pavani *et al.*, 2016; Mahdy *et al.*, 2017). According to Khodaei-Motlagh *et al.*, (2011), heat stress lowers blood progesterone levels, which is a major factor in aberrant oocyte maturation, implantation failure, and ultimately early embryonic demise in dairy cattle.

Effect of cortisol levels on conception, gestation and embryo development in cows

In cattle breeds managed under extensive system, high cortisol levels directly interfere with the physiological processes that control ovulation, conception and pregnancy establishment (Cooke, *et al.*, 2009; Faria *et al.*, 2023). Heat stress affects the expression of several genes linked to the growth and development of embryos including connexin 43 (Cx43), E-cadherin (CDH1), DNA methyltransferase 1 (DNMT1) and heat shock protein family A member 14 (HSPA14). The down-regulation of these genes may be the cause of the lower maturation rate. In contrast, the HSPA14 gene is a crucial component of the cell's machinery for folding, unfolding, transport, protein localization and differentiation, controlling the embryonic cell cycle and aiding in stress protection. As a result, HSPA14 which is an apoptotic gene brought on by heat shock is linked to the death of embryos and plays a significant part in the regulation of processes related to

growth, cellular differentiation and embryonic development (Pavani *et al.*, 2016). Additionally, Cx43 mediates the effects of heat stress on cells and is expressed in a variety of tissues including gonads (Mahdy *et al.*, 2017).

As an active glucocorticoid and anti-inflammatory hormone, cortisol modulates the synthesis and function of prostaglandins and cytokines necessary for ovulation, luteolysis, embryo implantation, foetal growth and placental development (Seth *et al.*, 2016). Cortisol can also modify corpus luteum function by affecting prostaglandin F2 α release (Duong *et al.*, 2012). Low rates of embryo implantation and consequently pregnancy loss are caused by cortisol's detrimental effects on the corpus luteum's progesterone release during the early stages of pregnancy (Parent *et al.*, 2002; Faria *et al.*, 2023). Research on temperament and acclimatization effect of stress on reproductive performance in beef cattle has shown that aggressive animals have higher plasma cortisol concentrations, lower rates of pregnancy, calving and weaning, as well as lower calf weight at birth and weaning (Cooke *et al.*, 2012). Excitable temperament also has a negative impact on pregnancy rates in cows subjected to natural breeding (Faria *et al.*, 2023).

Conclusion

Stress induced cortisol release has a significant and complex impact on the ability of cows to reproduce. Any stressor that activates the hypothalamic-pituitary-adrenal (HPA) axis or pathway and causes elevated cortisol levels in circulation has an inhibitory effect on reproduction or infertility in cows. Physiological and environmental stressors such as heat stress activate the HPA blocking the hypothalamic-pituitary-gonadal (HPG) axis suppressing the reproduction efficiency in cows. Follicular development, ovulation and oestrous expression are ultimately hampered

by this disturbance which results in decreased secretion of important reproductive hormones such as GnRH, LH, FSH, progesterone and oestradiol. Cortisol lowers oocyte quality and developmental competence at the ovarian level by adversely affecting folliculogenesis, oocyte maturation and steroidogenesis. Prolonged exposure to high cortisol concentrations also affects gene expression, impairs implantation and gestation processes thereby increasing embryonic mortality. Similarly, these physiological disruptions decrease conception rates, lower fertility and poor reproductive efficiency in cattle. Therefore, optimizing reproductive performance in cows requires effective management measures targeted at reducing stress, especially heat stress. A scientific foundation for boosting animal welfare, increasing productivity and creating focused interventions in cattle production systems is provided by comprehending the pathways through which cortisol disrupts reproductive processes.

Conflict of interest

The author declares that the research, authorship and publication of this review were all free from any conflicts of interest. There is no institutional, financial, personal or professional tie that might be interpreted as affecting the work's neutrality or integrity.

References

- Albiley TA, Johnson HD and Madam LM (1975). Influence of environmental heat on peripheral plasma progesterone and cortisol during the bovine oestrous cycle. *Journal of Animal Science*, 58: 1836 – 1840.
- Al-Katanani YM, Paula-Lopes FF and Hansen PJ (2002). Effect of season and exposure to heat stress on oocyte competence in

- Holstein cows. *Journal of Dairy Science*, 85: 390 – 396.
- Anbo N, Suzuki A, Mukangwa M, Takahashi R, Muranishi Y and Tetsuka M (2022). Progesterone stimulates cortisol production in the maturing bovine cumulus-oocyte complex. *Theriogenology* 189: 183 – 191.
- Andersen CY (2002). Possible new mechanism of cortisol action in female reproductive organs: physiological implications of the free hormone hypothesis. *Journal of Endocrinology*. 173(2): 211 – 217.
- Breen KM and Karsch FJ (2006). New insights regarding glucocorticoids, stress and gonadotropin suppression. *Frontiers in Neuroendocrinology*, 27: 233 – 245.
- Bridges PJ, Brusie MA and Fortune JE (2005). Elevated temperature (heat stress) in vitro reduces androstenedione and estradiol and increases progesterone secretion by follicular cells from bovine dominant follicles. *Domestic Animal Endocrinology*, 29(3): 508 – 522.
- Burke JM, Spiers DE, Kojima FN, Perry GA, Salfen BE, Wood SL, Patterson DJ, Smith MF, Lucy MC, Jackson WG and Piper EL (2001). Interaction of endophyte-infected fescue and heat stress on ovarian function in the beef heifer. *Biology of Reproduction*, 65: 260 – 268.
- Christison G and Johnson H (1972). Cortisol turnover in heat-stressed cows. *Journal of Animal Science Abstract*, 35(5): 1005 – 1010.
- Chrousos P, Torpy J and Gold W (1998). Interactions between the hypothalamic-pituitary-adrenal axis and the female reproductive system: clinical implications. *Annals of Internal Medicine* 129(3): 229 – 240.
- Collier RJ, Renquist BJ and Xiao Y (2017). A 100-Year Review: Stress physiology including heat stress. *Journal of Dairy Science*, 100: 10367 – 10380.
- Cooke RF, Arthington JD, Austin BR and Yelich JV (2009). Effects of acclimation to handling on performance, reproductive, and physiological responses of Brahman-crossbred heifers. *Journal of Animal Science*, 87: 3403 – 3412.
- Cooke RF., Bohnert DW, Cappellozza BI, Mueller CJ and Delcurto T (2012). Effects of temperament and acclimation to handling on reproductive performance of *Bos taurus* beef females. *Journal of Animal Science*, 90: 3547 – 3555.
- Coutinho AE and Chapman KE. (2011). The anti-inflammatory and immunosuppressive effects of glucocorticoids, recent developments and mechanistic insights. *Molecular Cell Endocrinology*, 335: 2 – 13.
- Das G, K and Khan FA (2010). Summer anoestrus in buffalo - A review. *Reproduction of Domestic Animal*, 45: e483 – e494.
- De Graaf Roelfsema E, Keizer A, Van E, Wijnberg D and Van H (2007). Hormonal responses to acute exercise, training and overtraining. A review with emphasis on the horse. *Veterinary Quarterly*, 29(3): 82 – 101.
- Dobson H and Smith F (1995). Stress and reproduction in farm animals. *Journal of Reproduction and Fertility*, 49: 451 – 461.
- Doney M, Gunn G and Griffiths G (1973). The effect of pre-mating stress on the onset of oestrus and on ovulation rate in Scottish blackface ewes. *Journal of Reproduction and Fertility*, 35(2): 381 – 384.
- Duong TH, Pitrowska KK, Bah MM, Sinderewicz E, Majewska M, Jankowska K, Okuda K and Skarzynski DJ (2012). Effects of cortisol on pregnancy rate and corpus luteum function in heifers: an in vivo study. *Journal of Reproduction and Development*, 58: 223 – 230.
- Elvinger F, Natzke RP and Hansen PJ (1992). Interactions of heat stress and bovine somatotropin affecting physiology and

- immunology of lactating cows. *Journal of Dairy Science*, 75: 449 – 462.
- Engler D, Pham T, Fullerton MJ, Ooi G, Funder JW and Clarke IJ (1989). Studies of the secretion of Corticotrophic Releasing Factor and Arginine vasopressin into the hypophyseal portal circulation of the conscious sheep. *Neuroendocrinology*, 49: 667 – 381.
- Faria N, Pacheco-Lima J, Moreira da Silva MH, Borba A and Moreira da Silva JF (2023). Effects of cortisol levels on reproductive success in cattle of different temperaments during fixed-time artificial insemination (FTAI). *Journal of Animal Behaviour and Biometeorology*, 11: e2023025.
<https://doi.org/10.31893/jabb.2024002>
- Fisher AD, Crowe MA, Alonso de la Varga ME and Enright WJ (1996). Effect of castration method and the provision of local anaesthesia on plasma cortisol, scrotal circumference, growth, and feed intake of bull calves. *Journal of Animal Science*, 74: 2336 – 2343.
- Gilad E, Meidan R, Berman A, Graber Y and Wolfenson D (1993). Effect of heat stress on tonic and GnRH-induced gonadotrophin secretion in relation to concentration of oestradiol in plasma of cyclic cows. *Journal of Reproduction and Fertility*, 99: 315 – 321.
- Gonzalez R, Ruiz Leon Y, Gomendio M and Roldan R (2010a). The effect of glucocorticoids on mouse oocyte *in vitro* maturation and subsequent fertilization and embryo development. *Toxicology In Vitro*, 24: 108 – 115.
- Gonzalez R, Ruiz Leon Y, Gomendio M and Roldan R (2010b). The effect of glucocorticoids on ERK-1/2 phosphorylation during maturation of lamb oocytes and their subsequent fertilization and cleavage ability *in vitro*. *Reproduction and Toxicology*, 29(2): 198 – 205.
- Hansen PJ (2007). Exploitation of genetic and physiological determinants of embryonic resistance to elevated temperature to improve embryonic survival in dairy cattle during heat stress. *Theriogenology*, 68(1): S242 – S249.
- Hapgood JP, Avenant C and Moliki JM (2016). Glucocorticoid-independent modulation of GR activity: Implications for immunotherapy. *Pharmacology and Therapeutics*, 165: 93 – 113.
- Heinz KG and Allrich RD (1992). Influence of exogenous adrenocorticotrophic hormone on oestrus behavior in cattle. *Journal of Animal Science*, 77: 735 – 739.
- Holmes MC and Seckl JR (2006). The role of 11beta-hydroxysteroid dehydrogenases in the brain. *Molecular Cell Endocrinology*, 248: 9 – 14.
- Howell JL, Fuquay JW and Smith AE (1994). Corpus luteum growth and function in lactating Holstein cows during spring and summer. *Journal of Dairy Science*, 77: 735 – 739.
- Hsueh J, Mc Gee A, Hayashi M and Hsu Y (2000). Hormonal regulation of early follicle development in the rat ovary. *Molecular Cell Endocrinology*, 163(1-2): 95 – 100.
- Kalin NH, Shelton SE and Turner JG (1992). Effects of b-carboline on fear-related behavioral and neurohormonal responses in infant rhesus monkeys. *Biological Psychiatry*, 31: 1008 – 1019.
- Khodaei-Motlagh M, Shahneh AZ, Masoumi R and Derensis F (2011). Alterations in reproductive hormones during heat stress in dairy cattle. *African Journal of Biotechnology*, 10(29): 5552 – 5558.
- Komesaroff A, Esler M, Clarke J, Fullerton J and Funder W (1998). Effects of estrogen and estrous cycle on glucocorticoid and catecholamine responses to stress in sheep. *American Journal of Physiology*, 275: E671 – E678.

- Krozowski Z, Li KX, Koyama K, Smith RE, Obeyesekere VR, Stein-Oakley A, Sasano H, Coulter C, Cole T and Sheppard KE (1999). The type I and type II 11 β -hydroxysteroid dehydrogenase enzymes. *Journal of Steroid Biochemistry and Molecular Biology*, 69: 391 – 401.
- Kumar P and Sharma A. (2014). Gonadotropin-releasing hormone analogs: Understanding advantages and limitations. *Journal of Human Reproductive Science*, 7(3): 170 – 174.
- Lee M, Ohmachi M, Arur S, Nayak S and Francis R (2007). Multiple Functions and Dynamic Activation of MPK-1 Extracellular Signal-Regulated Kinase Signaling in *Caenorhabditis elegans* Germline Development. *Genetics*, 177(4): 2039 – 2062.
- Ley SJ, Waterman AE and Livingston A (1996). Measurement of mechanical thresholds, plasma cortisol and catecholamines in control and lame cattle: A preliminary study. *Research in Veterinary Science*, 61: 172 – 173.
- Lotfi C and Mendonca P (2016). Comparative Effect of ACTH and Related Peptides on Proliferation and Growth of Rat Adrenal Gland. *Frontiers in Endocrinology* (Lausanne) 7: 39.
- Mahdy A, Pavani K, Baron E and Da Silva F (2017). Studies on Gene Expression and Developmental Competence of Bovine Embryos Produced Under Different Conditions of Heat Stress: Full Chapter. *Trends and Advances in Veterinary Genetics*. In Tech Open 3: 30 – 60.
- Martinez Miro S, Tecles F, Ramon M, Escribano D and Hernandez F (2016). Causes, consequences and biomarkers of stress in swine: an update. *BMC Veterinary Research*, 12: 171.
- Massey J, Campbell B, Raine Fenning N, Aujla K and Vedhara K (2014). The association of physiological cortisol and IVF treatment outcomes: a systematic. *Reproductive Medicine and Biology*, 13(4): 161 – 176.
- Michael AE, Thurston LM and Rae M T (2003). Glucocorticoid metabolism and reproduction: a tale of two enzymes. *Reproduction*, 126: 425 – 441.
- Minton JE (1994). Function of the hypothalamic-pituitary-adrenal axis and the sympathetic nervous system in models of acute stress in domestic farm animals. *Journal of Animal Science*, 72: 1891 – 1898.
- Molony V, Kent J and Robertson I (1993). Behavioural responses of lambs of three ages in the first three hours after three methods of castration and tail docking. *Research in Veterinary Science*, 55: 236 – 245.
- Mudron P, Rehage J, Sallmann H, Holtershinken M and Scholz H (2005). Stress response in dairy cows related to blood glucose. *Acta Veterinaria Brno*, 74(1): 37 – 42.
- Parent J, Chapdelaine P, Sirois J and Fortier MA (2002). Expression of microsomal prostaglandin E synthase in bovine endometrium: coexpression with cyclooxygenase type 2 and regulation by interferon-tau. *Endocrinology*, 143(8): 2936 – 2943.
- Pavani K, Baron E, Correia P, Lourenfo J, Bettencourt BF, Sousa M and Moreira da Silva F (2016) Gene expression, oocyte nuclear maturation and developmental competence of bovine oocytes and embryos produced after *in vivo* and *in vitro* heat shock. *Zygote*, 24(5): 748 – 759.
- Pavani K, Carvalhais I, Faheem M, Chaveiro A, Reis FV and Moreira da Silva F (2015). Reproductive performance of Holstein dairy cows grazing in dry- summer subtropical climatic conditions: Effect of heat stress and heat shock on meiotic competence and *in vitro* fertilization. *Asian-Australas Journal of Animal Science*, 28(3): 334 – 342.

- Pavani K, Rocha A, Baron E, Lourenfo J, Faheem M and Moreira da Silva F (2017). The effect of kinetic heat shock on bovine oocyte maturation and subsequent gene expression of targeted genes. *Zygote* 25(3): 383-389.
- Polsky L and Von Keyserlingk M (2017). Invited review: Effects of heat stress on dairy cattle welfare. *Journal of Dairy Science*, 100(11): 8645 – 8657.
- Prasad S, Tiwari M, Pandey A, Shrivastav T and Chaube S (2016). Impact of stress on oocyte quality and reproductive outcome. *Journal of Biomedical Science*, 23: 36.
- Raghavendra B, Sreenivasan Y and Manna K (2007). Oleandrin induces apoptosis in human, but not in murine cells: dephosphorylation of Akt, expression of FasL, and alteration of membrane fluidity. *Molecular Immunology*, 44(9): 2292 – 2302.
- Ralph CR, Lehman MN, Goodman RL and Tilbrook AJ (2016). Impact of psychosocial stress on gonadotrophins and sexual behaviour in females: role for cortisol? *Reproduction*, 152: R1 – R14.
- Ram S, Awasthi MK, Khan JR, Parveen K and Singh N (2023). Influence of cortisol level on ovarian follicular activity in postpartum Sahiwal cows during hot-humid season. *Haryana Vet*, 62(2): 6 – 9.
- Roman-Ponce H, Thatcher WW and Wilcox CJ (1981). Hormonal interrelationships and physiological responses of lactating dairy cows to a shade management system in a subtropical environment. *Theriogenology*, 16: 139 – 154.
- Scott E (2019). *Cortisol: The Stress Hormone*. Dotdash Publishing Family.
- Seckl JR (2004). 11 β -hydroxysteroid dehydrogenases: changing glucocorticoid action. *Current Opinions in Pharmacology*, 4: 597 – 602.
- Seth S, Lewis AJ and Galbally M (2016). Perinatal maternal depression and cortisol function in pregnancy and the postpartum period: a systematic literature review. *BMC Pregnancy and Childbirth*, 16(1): 124.
- Stilwell G (2012). Scientific opinion on the welfare of cattle kept for beef production and the welfare in intensive calf farming systems. *EFSA Journal* 10(5): 2669.
- Tetsuka M and Tanakadate M (2019). Activation of HSD11B1 in the bovine cumulus-oocyte complex during IVM and IVF. *Endocrinology Connect*, 8: 1029 – 1039.
- Tetsuka M, Takagi R, Ambo N, Myat TS, Zempo Y and Onuma A (2016). Glucocorticoid metabolism in the bovine cumulus-oocyte complex matured in vitro. *Reproduction*, 151: 73 – 82.
- Tilbrook AJ, Turner AI and Clarke IJ (2000). Effects of heat stress on reproduction in non-rodent mammals: the role of glucocorticoids and sex differences. *Journal of Reproduction and Fertility*, 5: 105 – 113.
- Timmermans S, Souffriau J and Libert CA (2019). General Introduction to glucocorticoid biology. *Frontiers in Immunology*, 10: 1545.
- Wade GN and Jones JE (2004). Neuroendocrinology of nutritional infertility. *American Journal of Physiology, Regulatory Integrative Comparative Physiology*, 287: R1277 – 1296.
- Wagenmaker E, Breen K, Oakley A, Pierce B, Tilbrook AJ, Turner AI and Karsch FJ (2009). Cortisol interferes with the estradiol-induced surge of luteinizing hormone in the ewe. *Biology of Reproduction*, 80(3): 458 – 463.
- Welsh R and Johnson H (1981). Stress-induced alterations in secretion of corticosteroids, progesterone, luteinizing hormone, and testosterone in bulls. *Endocrinology*, 109(1): 185 – 190.

- Wilson J, Marion S, Spain N, Spiers E, Keisler DH, and Lucy MC (1998) Effects of controlled heat stress on ovarian function of dairy cattle. 1. Lactating cows. *Journal of Dairy Science*, 81(8): 2124 – 2131.
- Wise ME, Armstrong DV, Huber JT, Hunter R and Wiersma F (1988). Hormonal alterations in the lactating dairy cow in response to thermal stress. *Journal of Dairy Science*, 71: 2480 – 2485.
- Wolfenson D, Roth Z and Meidan R (2000). Impaired reproduction in heat-stressed cattle: Basic and applied aspects. *Animal Reproductive Science*, 60-61: 535 – 547.
- Yuan H, Han X, He N, Wang G, Gong S, Lin J, Gao M and Tan JH (2016). Glucocorticoids impair oocyte developmental potential by triggering apoptosis of ovarian cells via activating the Fas system. *Scientific Report*, 6: 24036.